

An enzyme immunoassay (EIA) for the quantitative detection of AFP (Alpha-fetoprotein) in human serum or plasma.
For professional *in vitro* diagnostic use only.

INTENDED USE

The AFP EIA Test Kit is an enzyme immunoassay for *in vitro* quantitative determination of AFP level in human serum or plasma. It is intended as an aid in the assessment and diagnosis of fetal open neural tube defects and other types of cancers, such as ovarian, liver and testicular cancers.

SUMMARY

Alpha-Fetoprotein (AFP) is a glycoprotein with a molecular weight of approximately 70,000 daltons that shares sequence homology with albumin.^{1,2} AFP is normally produced during fetal and neonatal development by the liver, yolk sac, and small concentrations by the gastrointestinal tract.³ Fetal plasma AFP diffuses into the fetal urine and is excreted into the amniotic fluid from where it diffuses into the maternal circulation. The concentration of AFP in the fetal plasma peaks at 12 – 14 weeks and then rapidly falls.⁴ At birth, normal infants have AFP levels of above the normal range, but decreasing to within it over the first 1–2 years of life.⁵ By the second year of life, AFP concentrations decrease rapidly, and thereafter only trace amounts are normally detected in serum.⁶ In general, normal adults have serum AFP concentrations of less than 10 ng/mL. Elevated AFP levels occur in several malignant diseases including hepatocellular carcinoma, testicular nonseminomatous origin, and occasionally of other endodermal origin.⁷ Detection of elevated AFP levels can also be used in the detection of fetal open neural tube defects.⁸ The AFP EIA Test Kit is an immunoassay for the quantitative detection of the presence of Alpha-fetoprotein (AFP) in serum or plasma specimen. The test utilizes monoclonal antibodies to selectively detect AFP in serum or plasma.

PRINCIPLE

The AFP EIA Test Kit is a solid phase enzyme immunoassay based on a sandwich principle for the quantitative detection of AFP in human serum or plasma. The microwell plate is coated with monoclonal antibodies specific to AFP. During testing, the specimen and the enzyme-conjugated AFP antibodies are added to the antibody coated microwell plate and then incubated. If the specimen contains AFP, it will bind to the antibodies coated on the microwell plate and simultaneously bind to the conjugate to form immobilized antibody-AFP-conjugate complexes. If the specimen does not contain AFP, the complexes will not be formed. After initial incubation, the microwell plate is washed to remove unbound materials. Substrate is added and then incubated to produce a blue color, indicating the amount of AFP present in the specimen. Sulfuric acid solution is added to the microwell plate to stop the reaction which produces a color change from blue to yellow. The color intensity, which corresponds to the amount of AFP present in the specimen, is measured with a microplate reader at 450/630-700 nm or 450 nm. The absorbance of the specimen is then compared to a calibration curve to obtain the amount of AFP present in the specimen.

PRECAUTIONS

- For professional *in vitro* diagnostic use only. Do not use after expiration date.
- Do not mix reagents from other kits with different lot numbers.
- Avoid cross contamination between reagents to ensure valid test results.
- Add all the calibrators, controls, and specimens into the wells within 15 minutes to minimize the change in absorbance which may affect the results.
- Follow the wash procedure to ensure optimum assay performance.
- Use Plate Sealer to cover microwell plate during incubation to minimize evaporation.
- Use a new pipet tip for each specimen assayed.
- Ensure that the bottom of the plate is clean and dry and that no bubbles are present on the surface of the liquid before reading the plate. Do not allow wells to dry out during the assay procedure.
- Do not touch the bottom of the wells with pipette tips. Do not touch the bottom of the microwell plate with fingertips.
- Do not allow sodium hypochlorite fumes from chlorine bleach or other sources to contact the microwell plate during the assay as the color reaction may be inhibited.
- All equipment should be used with care, calibrated regularly and maintained following the equipment manufacturer's instructions.

HEALTH AND SAFETY INFORMATION

- Some components of this kit contain human blood derivatives. No known test method can offer complete assurance that products derived from human blood will not transmit infectious agents. Therefore, all blood derivatives should be considered potentially infectious. It is recommended that these reagents and human specimens be handled using established good laboratory working practices.
- Wear disposable gloves and other protective clothing such as laboratory coats and eye protection while handling kit reagents and specimens. Wash hands thoroughly when finished.
- ProCin™ 300 is included as a preservative in the Conjugate, Concentrated Wash Buffer, Substrate and Calibrators. Avoid any contact with skin or eyes.
- Do not eat, drink or smoke in the area where the specimens or kits are handled. Do not mouth pipette.
- Avoid any contact of the Substrate and Stop Solution with skin or mucosa. The Stop Solution contains 2M sulfuric acid which is a strong acid. If spills occur, wipe immediately with large amounts of water. If the acid contacts the skin or eyes, flush with large amounts of water and seek medical attention.
- Non-disposable apparatus should be sterilized after use. The preferred method is to autoclave for one hour at 121°C. Disposables should be autoclaved or incinerated. Do not autoclave materials containing sodium hypochlorite.
- Handle and dispose all specimens and materials used to perform the test as if they contained infectious agents. Observe established precautions against microbiological hazards throughout all the procedures and follow the standard procedures for proper disposal of specimens.
- Observe Good Laboratory Practices when handling chemicals and potentially infectious material. Discard all contaminated material, specimens and reagents of human origin after proper decontamination and by following local, state and federal regulations.
- Neutralized acids and other liquids should be decontaminated by adding sufficient volume of sodium hypochlorite to obtain a final concentration of at least 1.0%. A 30 minute exposure to a 1.0% sodium hypochlorite may be necessary to ensure effective decontamination.

STORAGE AND STABILITY

- Unopened test kits should be stored at 2-8°C upon receipt. All reagents are stable through the expiration date printed on the box. Return reagents to 2-8°C immediately after use.
- Allow the sealed pouch to reach room temperature before opening the pouch and removing the required number of strips to prevent condensation of the microwell plate. The remaining unused strips should be stored in the original resealable pouch at 2-8°C and can be used within 1 month of the opening date.

- Concentrated Wash Buffer may be stored at room temperature to avoid crystallization. If crystals are present, warm up the solution at 37°C. Working Wash Buffer is stable for 2 weeks at room temperature.
- Do not expose reagents especially the Substrate to strong light or hypochlorite fumes during storage or incubation steps.
- Do not store Stop Solution in a shallow dish or return it to the original bottle after use.

SPECIMEN COLLECTION AND PREPARATION

- The AFP EIA Test Kit can be performed using only human serum or plasma collected from venipuncture whole blood.
- EDTA, sodium heparin, and ACD collection tubes may be used to collect venipuncture whole blood and plasma specimens. The preservative sodium azide inactivates horseradish peroxidase and may lead to erroneous results.
- Separate serum or plasma from blood as soon as possible to avoid hemolysis. Grossly hemolytic, lipidic or turbid samples should not be used. Specimen with extensive particulate should be clarified by centrifugation prior to use. Do not use specimens with fibrin particles or contaminated with microbial growth.
- Do not leave specimens at room temperature for prolonged periods. Serum and plasma specimens may be stored at 2-8°C for up to 7 days prior to assaying. For long term storage, specimens should be kept frozen below -20°C.
- Bring specimens to room temperature prior to testing. Frozen specimens must be completely thawed and mixed well prior to testing. Specimens should not be frozen and thawed repeatedly.
- If specimens are to be shipped, they should be packed in compliance with local regulations covering the transportation of etiologic agents.

REAGENTS AND COMPONENTS

		Materials Provided			
No.	Reagent	Component Description	Quantity		
			96 wells/kit	480 wells/kit	48 wells/kit
	AFP Microwell Plate	Microwell plate coated with monoclonal Anti-AFP	1 plate (96 wells/plate)	5 plates (96 wells/plate)	1 plate (48 wells/plate)
1	AFP Conjugate	Anti-AFP bound to peroxidase; Preservative: 0.1% ProCin™ 300	1 x 12 mL	5 x 12 mL	1 x 6 mL
2	Concentrated Wash Buffer (25x)	Tris-HCl buffer containing 0.1% Tween 20; Preservative: 0.1% ProCin™ 300	1 x 40 mL	5 x 40 mL	1 x 20 mL
2A	Specimen Diluent	0.02M Phosphate buffered saline (PBS) buffer Preservative: 0.1% ProCin™ 300	1 x 12 mL	5 x 12 mL	1 x 6 mL
3	Substrate	Citrate-phosphate buffer containing hydrogen peroxide and tetramethylbenzidine (TMB); Preservative: 0.1% ProCin™ 300	1 x 12 mL	5 x 12 mL	1 x 6 mL
4	Stop Solution	2M Sulfuric acid	1 x 8 mL	5 x 8 mL	1 x 8 mL
5	AFP Calibrator 1	Diluted human serum non-reactive for AFP; Preservative: 0.1% ProCin™ 300	1 x 1 mL	5 x 1 mL	1 x 0.5 mL
6	AFP Calibrator 2	Diluted human serum containing 10 ng/mL AFP; Preservative: 0.1% ProCin™ 300	1 x 1 mL	5 x 1 mL	1 x 0.5 mL
7	AFP Calibrator 3	Diluted human serum containing 20 ng/mL AFP; Preservative: 0.1% ProCin™ 300	1 x 1 mL	5 x 1 mL	1 x 0.5 mL
8	AFP Calibrator 4	Diluted human serum containing 50 ng/mL AFP; Preservative: 0.1% ProCin™ 300	1 x 1 mL	5 x 1 mL	1 x 0.5 mL
9	AFP Calibrator 5	Diluted human serum containing 100 ng/mL AFP; Preservative: 0.1% ProCin™ 300	1 x 1 mL	5 x 1 mL	1 x 0.5 mL
10	AFP Calibrator 6	Diluted human serum containing 400 ng/mL AFP; Preservative: 0.1% ProCin™ 300	1 x 1 mL	5 x 1 mL	1 x 0.5 mL
	Plate Sealers		2	10	2
	Package Insert		1	1	1

Materials Required But Not Provided

- Freshly distilled or deionized water
- Sodium hypochlorite solution for decontamination
- Absorbent paper or paper towel
- Water bath or incubator capable of maintaining 20°C to 30°C.
- Calibrated automatic or manual microwell plate washer capable of aspirating and dispensing 350 µL/well
- Disposable gloves
- Automated processor (optional)
- Calibrated micropipettes with disposable tips capable of dispensing 25, 50 and 100 µL
- Graduated cylinders for wash buffer dilution
- Vortex mixer for specimen mixing (optional)
- Disposable reagent reservoirs
- Calibrated microplate reader capable of reading at 450 nm with a 630-700 nm reference filter, or reading at 450 nm without a reference filter
- Timer

DIRECTIONS FOR USE

Allow reagents and specimens to reach room temperature (15-30°C) prior to testing. The procedure must be strictly followed. Assay must proceed to completion within time limits. Arrange the calibrators in a horizontal or vertical configuration. The procedure below assigns specific wells arranged in a vertical configuration. Configuration may depend upon software.

Step	Detailed Procedure	Simplified Procedure
	<ul style="list-style-type: none"> Prepare Working Wash Buffer by diluting the Concentrated Wash Buffer 1:25. Pour the contents of the bottle in a graduated cylinder and fill it with freshly distilled or deionized water to 1000 mL. It is stable for 2 weeks at 15-30°C. Note: If crystals are present in the Concentrated Wash Buffer, warm it up at 37°C until all crystals dissolve. 	<ul style="list-style-type: none"> Prepare Working Wash Buffer by diluting the Concentrated Wash Buffer 1:25
0	<ul style="list-style-type: none"> Leave A1 as Blank well. 	<ul style="list-style-type: none"> Leave A1 as Blank well
1	<ul style="list-style-type: none"> Add 25 µL of Calibrator 1 in wells B1 and C1. Add 25 µL of Calibrator 2 in wells D1 and E1. Add 25 µL of Calibrator 3 in wells F1 and G1 Add 25 µL of Calibrator 4 in wells H1 and A2. Add 25 µL of Calibrator 5 in wells B2 and C2 Add 25 µL of Calibrator 6 in wells D2 and E2. <p>The colors of Calibrator 1-6 gradually change from yellow to blue.</p>	<ul style="list-style-type: none"> B1and C1: Add 25 µL Calibrator 1 D1and E1: Add 25 µL Calibrator 2 F1and G1: Add 25 µL Calibrator 3 H1and A2: Add 25 µL Calibrator 4 B2and C2: Add 25 µL Calibrator 5 D2and E2: Add 25 µL Calibrator 6

2	<ul style="list-style-type: none"> Add 25 µL of specimen to assigned wells starting at F2. Remove unused strips from the microwell plate, and store in the original resealable pouch at 2-8°C. 	<ul style="list-style-type: none"> Starting F2: Add 25 µL specimen Remove and store unused strips at 2-8°C.
3	<ul style="list-style-type: none"> Add 100 µL of Conjugate to each well. (Red reagent) 	<ul style="list-style-type: none"> Add 100 µL of Conjugate to each well
4	<ul style="list-style-type: none"> Mix gently by swirling the microwell plate on a flat bench for 30 seconds. Cover the microwell plate with the Plate Sealer, and incubate at room temperature (20-30°C), in a water bath or in an incubator at 20-30°C for 30 minutes ± 5 minute. 	<ul style="list-style-type: none"> Mix gently Cover the microwell plate with the Plate Sealer and incubate at room temperature (20-30°C) for 30 min
5	<ul style="list-style-type: none"> Remove the Plate Sealer. Wash each well 5 times with 350 µL of Working Wash Buffer per well, then remove the liquid. Turn the microwell plate upside down on absorbent tissue for a few seconds. Ensure that all wells have been completely washed and dried. Note: Inproper washing may cause false positive results 	<ul style="list-style-type: none"> Remove the Plate Sealer Wash each well 5 times with 350 µL of Working Wash Buffer Turn the microwell plate upside down on absorbent tissue
6	<ul style="list-style-type: none"> Add 100 µL of Substrate to each well. (Clear Reagent) Then a light blue to blue color should develop in wells corresponding to the amount of AFP present in the specimen. 	<ul style="list-style-type: none"> Add 100 µL of Substrate to each well
7	<ul style="list-style-type: none"> Mix gently then cover microwell plate with Plate Sealer, and incubate at room temperature (20-30°C), in a water bath or in an incubator at 20-30°C for 15 minutes ± 2 minutes. 	<ul style="list-style-type: none"> Mix then cover microwell plate with Plate Sealer and incubate at room temperature (20-30°C) for 15 min
8	<ul style="list-style-type: none"> Remove the Plate Sealer. Add 50 µL of Stop Solution to each well. (Clear Reagent) Then a yellow color should develop in wells containing positive specimens Then a light yellow to yellow color should develop in wells corresponding to the amount of AFP present in the specimen. 	<ul style="list-style-type: none"> Remove Plate Sealer Add 50 µL of Stop Solution to each well
9	<ul style="list-style-type: none"> Read at 450/630-700 nm within 30 minutes. Note: Microwell plate can also be read at 450 nm, but it is strongly recommended to read it at 450/630-700 nm for better results. 	<ul style="list-style-type: none"> Read at 450/630-700 nm within 30 min

AUTOMATED PROCESSING

Automatic EIA microplate processors may be used to perform the assay after validating the results to ensure they are equivalent to those obtained using the manual method for the same specimens. Incubation times may vary depending on the processors used but do not program less incubation times than the procedure listed above. When automatic EIA microplate processors are used, periodic validation is recommended to ensure proper results.

QUALITY CONTROL

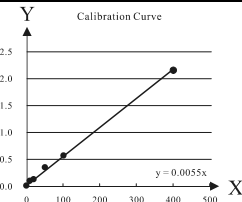
Control standards are not supplied with this kit; however, it is recommended that normal, low and high controls be tested with each run as a good laboratory practice to monitor assay performance. Each laboratory should establish its own criteria for establishing mean values and acceptable ranges to determine reliability of the results.

CALCULATION OF RESULTS

Draw the calibration curve and obtain quantitative specimen results.

- Calculate the Mean Absorbance of each Calibrator, then plot them on the Y-axis against their concentration on the X-axis on a linear graph-fitted and draw the calibration curve. Draw the best-fitted line through data points and zero point to obtain a standard curve. Refer to an example of the calibration curve at right.

NOTE: Do not use the calibration curve at right to make any calculation. A calibration curve must be performed for each run.



Exampe of Specimen Result Calculation

Item	Well	Absorbance	Mean (Absorbance – Blank)	Concentration (ng/mL)
Unknown Specimen	F2	0.445	0.437	79.45

- Obtain quantitative specimen results of concentrations expressed in ng/mL, from their absorbance by using the calibration curve. When the alternate result unit, IU/mL, is selected, the conversion factor used by the system is by multiplying by 0.83.

NOTE: Specimens that have absorbance above Calibrator 6 should be pre-diluted using 0.02M PBS and retested. The concentration must be multiplied by the dilution factor. Automated reading and calculation may also be performed using linear regression function on suitable computer programs.

LIMITATIONS

- The AFP EIA Test Kit is used for the detection of AFP in human serum or plasma. Diagnosis should not be established based on a single test result. Further testing should be performed in assessing clinical status. Specimens containing precipitate may give inconsistent test results.
- As with all diagnostic tests, all results must be interpreted together with other clinical information available to the physician.
- As with other sensitive immunoassays, there is the possibility that the positive result cannot be repeated due to inadequate washing from the initial test. The results may be affected due to procedural or instrument error.
- Unusually high titers of heterophilic antibodies or rheumatoid factor (RF) may affect results. Even if test results are positive, further clinical evaluation should be considered with other clinical information available to the physician.

EXPECTED VALUES

It is recommended that each laboratory establish its own range of expected values based on patient populations. A study to determine expected values using the AFP EIA Test Kit was conducted for initial reference use only.

Population	No. Specimens	0-10.0 ng/mL	10-20 ng/mL	>20ng/mL
Normal	982	98.88%	0.92%	0.20%

PERFORMANCE CHARACTERISTICS

Analytical Sensitivity

The analytical sensitivity of the AFP EIA Test Kit is 0.6 ng/mL using the standard procedure.

Accuracy

The AFP EIA Test Kit has been compared to a leading commercial AFP EIA test using clinical specimens. A total of 654 clinical specimens ranging from 20-400 ng/mL were run and analyzed using least square regression analysis. The results show that the AFP EIA Test Kit has good correlation compared to the reference method.

Reproducibility

Intra-Assay: Within-run precision has been determined by using 60 replicates of two specimens: a low positive and a medium positive.

Inter-Assay: Between-run precision has been determined by 180 independent assays on the same two specimens: a low positive and a medium positive. Three different lots of the AFP EIA Test Kit have been tested using these specimens over a 3-day period.

Specimen	Intra-Assay			Inter-Assay		
	Mean AFP Concentration (ng/mL)	Standard Deviation	Coefficient of Variation (%)	Mean AFP Concentration (ng/mL)	Standard Deviation	Coefficient of Variation (%)
1	24.950	2.289	9.173	24.581	2.336	9.504
2	225.308	14.668	6.510	224.367	15.175	6.763

Recovery and Linearity

Recovery: Known amounts of AFP were added to normal human serum with endogenous AFP concentration of 1.56 ng/mL. The concentration of AFP was determined using AFP EIA Test Kit and the resulting percent recovery was calculated.

Specimen	AFP Concentration Added (ng/mL)	AFP Concentration Obtained (ng/mL)	Recovery* (%)
Level 1	20.15	22.61	104.47
Level 2	40.26	40.95	97.84
Level 3	85.67	92.77	106.47
Level 4	170.48	180.85	105.17
Level 5	338.90	333.41	97.92

* Recovery = (Concentration Obtained (ng/mL) – Endogenous Level (ng/mL))/Concentration Added (ng/mL)

Linearity: Specimens containing known concentration of AFP were diluted with normal human serum and determined. The obtained concentrations were within ± 20% of the expected values.

Cross-Reactivity

The specificity of the AFP EIA Test Kit was determined by testing sera containing the compounds listed below. These compounds showed less than 20% interference in the AFP EIA Test Kit at the levels indicated.

Substance	Concentration	Substance	Concentration
Uric cid	0.09 mg/mL	Bilirubin	0.15 mg/mL
Vitamin	19.80 mg/mL	EDTA	0.20 mg/mL
Globin	0.99 mg/mL	Hemoglobin	16.67 mg/mL
Genitistic cid	0.20 mg/mL	Creatin	0.20 mg/mL
Acetaminophen	0.20 mg/mL	Cyclophosphamide	0.50 mg/mL
Oxalic cid	0.99 mg/mL	5-fluorouracil	2.00 mg/mL
Albumin	40.00 mg/mL	Cytosine arabinoside	0.30 mg/mL
Coffein	0.10 mg/mL		

Dose Hook Effect

No dose hook effect is observed up to 10,000 ng/mL of AFP.

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Index of Symbols

	Attention, see instructions for use		Tests per kit		Manufacturer
	For <i>in vitro</i> diagnostic use only		Use by		Catalog #
	Store between 2-8°C		Lot Number		Specimen Diluent
	AFP		Substrate		Conjugate
	Wash Buffer (25x)		Stop Solution		Calibrator 3
	Calibrator 1		Calibrator 2		Calibrator 6
	Calibrator 4		Plate Sealer		Package Insert
	Microwell Plate				